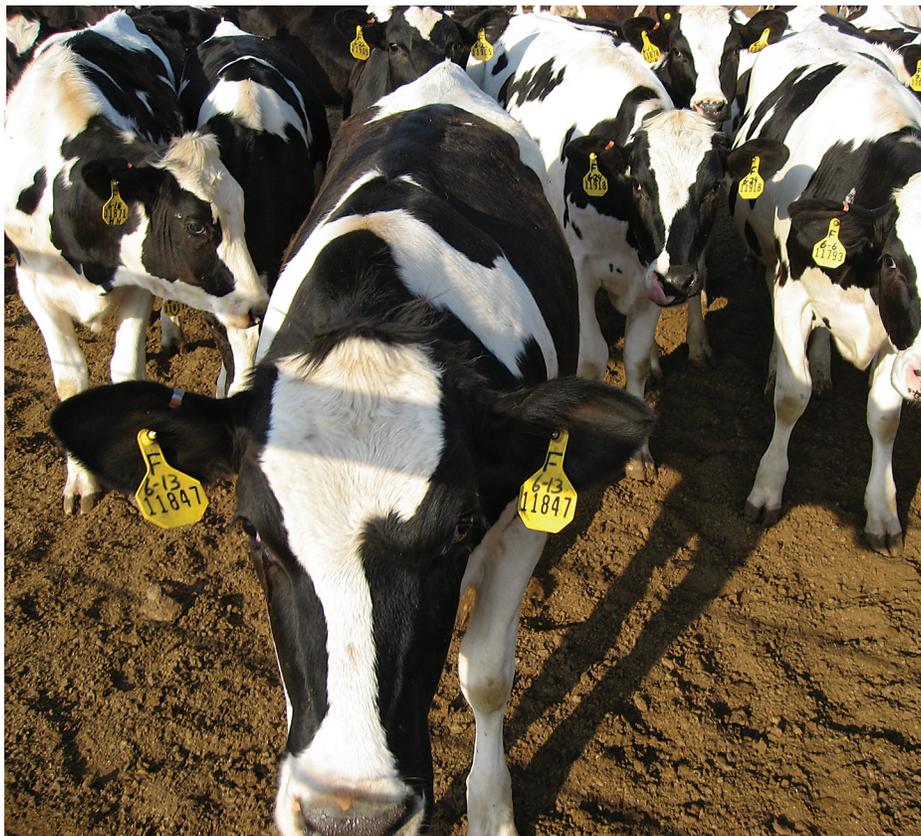


Testing new dairy cattle for disease can boost herd health, cut costs

by Dale A. Moore, John M. Adaska,
Gerald E. Higginbotham, Alejandro R. Castillo,
Carol Collar and William M. Sischo

Dairy producers seldom test or examine incoming cattle, although these important biosecurity practices are recommended. This pilot project examined risk management decisions that producers make when faced with test-positive animals in purchased groups of dairy cattle, in order to provide information on disease risks and conditions that could affect animal health and performance. New arrivals to seven herds at dairy farms in four California counties were examined and tested for a range of conditions. The most common findings were bovine leukosis virus (33% of cattle purchased) and male reproductive abnormalities (16% of bulls purchased). Once testing results were known, producers made a variety of risk management decisions. Although testing costs for some conditions outweigh the benefits of finding an infected animal, an individual producer's decision to test new animals most likely depends on their knowledge of the pros and cons as well as their risk tolerance.

The farm is considered the front line of food system security, and biosecurity practices such as disease testing are the primary means to protect the farm. Attention to on-farm biosecurity for livestock has been spurred by current certification and eradication programs for Johne's disease and bovine viral diarrhea virus (BVDV), the foot-and-mouth-disease outbreak in British sheep and cattle in 2001, and the letter-borne anthrax bioterrorism attacks in 2001 (Sandvik 2004; Sockett 1996; US Senate 2006).



Every year, California milk producers import about 120,000 to 130,000 head of dairy cattle into the state, primarily heifers. Yet surveys have found that the vast majority of producers do not test incoming animals for economically important conditions and communicable diseases.

The dairy industry in the United States has not widely adopted biosecurity practices, particularly those related to purchased cattle. In Wisconsin, less than 50% of producers with recently purchased cattle asked about the herd of origin's disease status, and less than 20% did any testing of animals they purchased (Hoe and Ruegg 2006). In Idaho, 80% of herds undergoing expansion did not require health testing for new cattle, except for mastitis detection (Dalton et al. 2005a). In addition, only about 40% of producers purchasing bulls quarantined them on arrival and only about 25% required a breeding soundness examination (Dalton et al. 2005b). In the upper Midwest, nearly 60% of herds undergoing expansion obtained cattle with minimal health histories, and less than half required any health testing (Faust et al. 2001). Yet owners and managers involved in herd expansions indicated that herd health

was compromised as a result of expansion. Similarly, Canadian farms that purchased replacement animals had more cattle testing positive for Johne's disease than farms that did not purchase animals (Chi et al. 2002).

There are approximately 1.7 million head of dairy cattle in California, which produced about 41 billion pounds of milk in 2007, generating an estimated \$61.4 billion in economic activity in the state. The movement of dairy cattle into California is a risk to the state's herds. The state imports approximately 120,000 to 130,000 head of dairy cattle annually from other states and countries, a rate of about 10,000 to 13,000 animals per month. Although this includes mature cows and bulls, the trade is primarily in Holstein and Jersey heifers or young stock (6 to 24 months old). Each year, California imports heifers from as many as 35 states and from as far away as New Hampshire (M. Ashcroft,

California Department of Food and Agriculture Animal Health Branch, personal communication, March 2006).

Dairy farm biosecurity

Good management practices for biosecurity focus on efforts to prevent the entry of diseases onto the farm as well as to prevent disease transmission within the farm (see box) (Buhman et al. 2000). An important element is the pre-purchase evaluation of cattle, because standard diagnostic laboratory tests or clinical examinations can detect many agents associated with clinical disease outbreaks that could

be economically significant. These diseases include: bovine viral diarrhea virus (BVDV) persistent infection, which can spread from animal to animal and cause abortions or congenital defects; *Salmonella*, which can cause diarrhea in adult animals and young stock; mastitis or udder infections caused by *Mycoplasma bovis*, *Staphylococcus aureus* and *Streptococcus agalactiae*; and digital dermatitis, or foot warts, which causes lameness. Other diseases, such as bovine leukosis virus (BLV, a cancer-producing retrovirus) infections and Johne's disease, which can cause diarrhea and production losses, are insidious and so do not manifest disease in outbreak form. However, although insidious diseases initially have few if any detectable symptoms, they can eventually cause clinical disease and affect the marketability of animals. One of the diseases, *Salmonella*, can also infect people. Examining cattle and testing them for endemic diseases or other abnormalities upon arrival to the farm will not prevent the entry of all diseases, but it is the first step to reducing their introduction and provides a screening mechanism for diseases that could result in an epidemic.

Efforts by Cooperative Extension, animal agriculture organizations and others to educate cattle producers about biosecurity are extensive and include all the important disease prevention strategies. Nonetheless, many producers have not yet adopted testing for new herd additions. To better understand the decisions producers make about the testing and disposition of test-positive animals, we did a pilot project

to develop a protocol for testing purchased cattle, in collaboration with UC Cooperative Extension (UCCE), herd veterinarians, Agricultural Experiment Station university scientists and the California Animal Health and Food Safety Laboratory.

Pre-purchase survey

Dairy farms enrolled in the study came from four California counties (Fresno, Kings, Stanislaus and Tulare) and were a convenience sample (not randomly selected) selected by UCCE farm advisors and practicing veterinarians. Eligible producers had to be actively engaged in purchasing animals and expect to purchase animals within 30 days of agreeing to participate in the survey. The herd owner completed a pre-purchase survey provided by the extension advisor or herd veterinarian. Survey questions focused on cattle purchased in the previous year, purchasing practices, disease testing, the examination of purchased cattle and the disposition of animals with specific disease conditions.

Seven dairy herds were enrolled, and all producers had purchased lactating animals the previous year. Five of the seven producers brought in new bulls and three bought bred (pregnant) heifers. None had information about specific disease history. Five of the producers knew the herd of origin but no testing was done. The exceptions were one producer who tested for Johne's disease, and four producers who checked for foot warts (digital dermatitis). Of the five farms purchasing bulls, two had breeding soundness exams done.

Biosecurity practices recommended for cattle premises

- Know the health history of herds from which cattle are purchased.
- Know the health status of animals purchased or brought into the operation.
- Request that the herd veterinarian talk to the seller's veterinarian prior to purchasing animals.
- Never purchase unvaccinated animals.
- Never buy animals from a herd that has mixed-origin cattle.
- Transport animals in clean vehicles.
- Have a control program for outside animals that could spread disease (rodents, etc.).
- Load and unload animals and supplies in areas located at the perimeter of the operation.
- Provide an isolated pickup area for rendering trucks to pick up mortality, to prevent contamination of the operation.
- Limit the number of visitors who have access to cattle pens, feed mixing and storage areas, and treatment areas.
- Keep a record of visitors to the operation.

Adapted from Buhman et al. 2000.

TABLE 1. Prevalence of test-positive animals in pilot study of newly purchased arrivals to California dairy farms

Condition	Animals tested	Test-positive
	no.	no. (%)
Bovine leukosis virus (BLV)	382	127 (33)
Bovine viral diarrhea virus (BVDV) persistent infection	382	0 (0)
Johne's disease	382	1 (0.26)
<i>Salmonella</i> spp.*	380	2 (0.53)
<i>Mycoplasma</i> spp. intramammary infection	373	0 (0)
<i>Staph. aureus</i> intramammary infection	373	3 (0.8)
<i>Strep. agalactiae</i> intramammary infection	373	0 (0)
Environmental <i>Staph.</i> spp.	373	10 (2.7)
Environmental <i>Strep.</i> spp.	373	8 (2.1)
Other intramammary infections	373	3 (0.8)
Abnormal bull genital findings	38	6 (15.8)

* *Salmonella* St. Paul and Mbandaka.

Five of the seven producers would not have purchased test-positive animals if they had known they were infected.

Five producers reported that they never isolate purchased animals upon arrival to the farm. Five producers did not cull animals positive for BVDV persistent infection or John's disease, but four usually culled cows positive for *S. aureus* mastitis.

Testing newly purchased cattle

The sample size for the number of cattle to be tested was based on an estimate of 120,000 animals purchased annually in California per 1.7 million head in the state dairy herd, or about 7.0% (CDFA 2005). If owners of the approximately 820,000 dairy cattle in the four-county study area reflect state trends, they purchase approximately 54,000 cattle per year. Detecting a 1.0% prevalence of the targeted diseases in these newly purchased cattle with 95% confidence would require a sample size of 298 animals. An extra 25% was added in case some cattle could not be found or subsequently tested, making a total of 372 animals to test and examine in the four-county area.

All new arrivals were examined by the project investigation team or herd veterinarian within 7 days of arrival. All cows, bulls and heifers received a general physical, and bulls also received a palpation examination of scrotal contents and seminal vesicles. A blood sample was obtained for the following infectious disease tests: BVDV antigen-capture ELISA (enzyme-linked immunosorbent assay), BLV antibody ELISA and, for cattle over 2 years of age, John's disease antibody-capture ELISA. Blood samples were processed by the California Animal Health and Food Safety Laboratory in Tulare. Milk samples from each quarter were obtained from all cows and post-calving heifers, and evaluated for common contagious mastitis pathogens (including *S. aureus*, *S. agalactiae* and *Mycoplasma* spp.) and for "environmental" pathogens like *E. coli* and *Streptococcus* species, by the UC Davis Veterinary Medicine Teaching and Research



Veterinarians examined 382 newly purchased animals in seven herds. The most common finding was bovine leukosis virus (33% of cattle purchased). Three of the seven producers surveyed decided to keep BLV-positive animals in their herds, but four out of seven would not have purchased the infected animals if they had known.

Center's Milk Quality Laboratory in Tulare. Fecal samples were evaluated for the presence of *Salmonella*.

Within the first week of arrival to participating farms, 382 dairy cattle were examined and tested. Of these, 25% were pre-calving heifers, 65% were lactating cows and 10% were bulls. Most of the cattle (72.8%) came from private owner sales and the rest through a sales yard, cattle buyer or auction. In addition, 57% of the cattle had U.S. Department of Agriculture (USDA) ear tags placed in California, 21% had ear tags from other states (Hawaii, Minnesota, North Dakota, South Dakota, Colorado, Washington and Oregon) and 22% had no official USDA ear tag.

The most common finding was evidence of BLV infection (33%) (table 1). Only one bull was BLV test-positive but over 35% of the cows and heifers were, making them 13 times more likely to test positive for this disease than bulls ($P < 0.001$). The proportion of BLV-positive purchased animals varied by destination farm, and ranged from 7% to 80%. Of 38 bulls evaluated, one was cryptorchid (right testicle not descended) and five had firm swellings of

either a seminal vesicle or epididymus, parts of the male reproductive tract. Swellings in these structures can indicate previous or current inflammation that could impair fertility.

Producer reactions to tests

Participating producers received a standard report of the physical examination findings and laboratory results. A questionnaire, provided within 2 weeks of sampling, captured producer decisions for each test-positive animal: (1) marked the cow/heifer with, for example, a leg band and kept her in the herd; (2) kept her but moved or will move her to a separate string for cows with that kind of infection; (3) removed the cow/heifer from the herd; (4) treated the cow/heifer; or (5) have not yet decided what to do with the animal(s). The questionnaire also asked producers whether they would have purchased the animal had they known the test results beforehand.

When provided with test-positive results, most producers indicated that they would keep the animals in the herds rather than cull but would not have purchased the cow had they known that she had the disease. For the

TABLE 2. Decisions by dairy producers after receiving test-positive results for intramammary infections

Dairy	Intramammary infection (no. cows)	What was done with animal(s)?	If knew animal infected before purchase	Decisions about infections
1	<i>Strep. spp.</i> (5) <i>Staph. spp.</i> (3) <i>Staph. aureus</i> (3) <i>Corynebacterium</i> (1)	Nothing	Would not have purchased	Ask veterinarian
2	<i>Strep. spp.</i> (1) <i>Staph. spp.</i> (2) <i>Corynebacterium</i> (1)	Kept but moved to a separate string for cows with same infection	Bought but asked for price discount	Make own decision
6	<i>Strep. spp.</i> (1) <i>Staph. spp.</i> (2)	Treated with intramammary antibiotics	Would not have purchased	Make own decision
7	<i>Strep. spp.</i> (1) <i>Staph. spp.</i> (3)	Treated with intramammary antibiotics	Would not have purchased	Make own decision

cow with suspected Johne’s disease, the producer said he would make his own decision about what to do with her. For intramammary infections, the producers made the same decision for each of their positive cattle, regardless of the type of bacteria found (table 2). For the *Salmonella* St. Paul–positive cow, the producer said he would ask his veterinarian what to do. For the *Salmonella* Mbandaka–positive cow, the producer said he would make his own decision about what to do.

Decisions regarding BLV-positive animals were producer-specific. Each producer reported making the same decision for each of the animals testing positive for any condition in their herds (table 3). Five of the seven producers would not have purchased test-positive animals if they had known they were infected. The producer with affected bulls decided to remove all these animals from his herd, would not have purchased them knowing they were affected, and would make his own decisions about what to do with them.

Making biosecurity decisions

In this pilot study, decision-making by dairy producers varied. Knowing infection status before purchasing can provide information for decisions about treatment, isolation or culling, but the

participating producers had different levels of risk-tolerance and said they would seek veterinary advice to varying degrees. Even though these producers were regularly purchasing animals and may have had infectious diseases in their herds in the past, these factors did not appear to influence their decision to require testing. In addition, new cattle were rarely isolated or quarantined: only two of the seven producers isolated some new additions on arrival, confirming the results of previous surveys (Buttars et al. 2006; Dalton et al. 2005a; Faust et al. 2001; Hoe and Ruegg 2006).

The perception of risk among farmers does not always translate into risk-tolerant or risk-averse behavior. In a study of swine producers, the perceived importance of a biosecurity practice was not necessarily associated with its implementation (Casal et al. 2007). The swine producers were more likely to implement biosecurity measures that affected disease transmission through people and wildlife than to implement measures for the most important risk for disease transmission: incoming replacement animals.

Costs and benefits of testing

The cost of examination and testing is a likely deterrent to producers purchasing large groups of animals. However,

several results of this pilot project indicate that there is some value in examining and testing for certain conditions.

Intramammary infections. If the farm strategy is to keep *S. aureus* intramammary infections out of a herd and the probability of infection is 0.8%, the cost of milk cultures to find one infected cow in 100 is about \$625. This is less than the cost of the average purchased cow minus her salvage value if she is tested after purchase (about \$1,800 and \$400 to \$500, respectively). If the bacteria spreads and infects other cows, the costs due to clinical mastitis could average about 726 pounds (330 kilograms) of milk per lactation, about \$120 (Shim et al. 2004).

Bovine leukosis virus. Evidence of BLV infection was the most common finding in our study. The consequences of BLV infection can include immunosuppression, premature culling, loss of salvage value if the animal becomes clinical and is culled, higher calving intervals and lower milk production (D’Angelino et al. 1998; Ott et al. 2003; Pollari et al. 1992, 1993). A few studies have found no influence of BLV infection on herd performance (Heald et al. 1992; Tiwari et al. 2007). However, a recent study that controlled for other factors associated with milk production found a significant relationship

TABLE 3. Decisions by dairy producers after receiving test-positive results for bovine leukosis virus (BLV) infections

Dairy	Total purchased	Tested BLV-positive	What was done with animal(s)?	If knew animal infected before purchase	Decisions about infections
	no.	no. (%)			
1	61	22 (36.1)	Kept in herd	Other	Ask veterinarian
2	20	16 (80)	Kept in herd	Would have bought anyway	Make own decision
3	6	3 (50)	No response	No response	No response
4	83	15 (18.1)	Removed animals	Would not have purchased	Make own decision
5	14	1 (7)	Kept in herd	Would not have purchased	Ask veterinarian
6	80	15 (18.8)	Not yet decided	Would not have purchased	Ask veterinarian
7	118	55 (46.6)	Not yet decided	Would not have purchased	Ask veterinarian

between higher herd prevalence of BLV and lower milk production and annual value of production (a combination of milk production and annual value of calves at birth, minus the annual net replacement cost) (Ott et al. 2003). Given Ott's model, a herd with a BLV prevalence of 33% has 253 pounds (115 kilograms) less milk per cow in the herd (1% lower production compared to cows in herds without BLV).

BLV infects lymphocytes, resulting in a lifelong infection, and can result in lymphosarcoma or malignant lymphoma. If 0.1% to 5.0% of BLV-positive animals develop lymphosarcoma and the herd prevalence of BLV is 33%, as many as 1% of animals in the herd will be culled prematurely due to the development of lymphosarcoma (Pelzer 1997). Premature culling incurs losses due to the replacement of a cow with a heifer, loss of pregnant cows and loss of the cow's market value (Rhodes et al. 2003). Nationwide, 5,175,861 beef and dairy cows were sent to market in 2002 (USDA 2008). Of those, 2.77% (143,484) were condemned, and 17% (25,075) of those condemnations were for malignant lymphoma, resulting in no value to the producer or packer.

The laboratory cost for a BLV test is about \$8.70 for 10 samples. In the case of BLV infections from purchased cattle, the risk is real and the potential consequences significant. Thus, BLV test results should be considered in dairy-cattle purchasing decisions.

Johne's disease. Only one Johne's disease test-positive animal was found in our study, resulting in less than 1% prevalence. However, the sensitivity of tests for this disease is notoriously low (Collins et al. 2006). The ELISA test on serum has a sensitivity of about 30% to 50%, which is the probability that a test is positive given that the animal is truly infected (Collins et al. 2006). As such, a negative Johne's disease test does not necessarily mean "not infected." Collins et al. (2006) provided a cattle purchase flowchart for Johne's disease biosecurity, which showed that the highest risk for buying infected cattle is from untested herd replacements.

The costs of Johne's-positive herds include reduced milk yield, body weight losses, a reduction in market cow beef value and early culling. About

\$60 to \$90 of income per cow is lost in Johne's-positive herds compared to negative herds (Collins et al. 2006; Ott et al. 1999). The prevalence of Johne's-infected dairy cattle is estimated to be about 22% nationwide, but it varies by region (table 4) (Ott et al. 1999). Although the West appears to have a relatively low proportion of Johne's-positive herds, buying replacement animals can put Western dairies at risk for introducing the disease. Laboratory tests for this disease are about \$3.60 for one and \$5.50 for 10 samples.

Bull diseases. Replacement bulls can bring in disease as well as be poor performers. If it costs \$50 to sample, test and conduct a 5-minute reproductive exam on a single bull, the 38 bulls in our project represented a total testing cost of \$1,900, or about the value of one bull. With one cryptorchid bull and five bulls with evidence of reproductive tract problems that could affect fertility, the testing cost can be justified. Adding a test (about \$8 each) for trichomoniasis, a disease spread venereally that can affect cow fertility, would also be an important biosecurity measure.

Testing as an insurance policy

Decisions to test cattle purchases for infectious diseases depend on both the risk of disease introduction and the risk aversion (or tolerance) level of each producer. Just as with making a decision about purchasing an insurance policy, individuals decide whether they can absorb the costs of some negative event or if they want to minimize risks associated with infectious diseases. Producers who decide not to test incoming cattle can employ three other strategies: (1) ask about the herd of origin and any disease information the sellers have, which will still not address carrier animals; (2) carefully examine cattle, particularly breeding bulls, to detect any obvious abnormalities before purchasing; and (3) provide an isolation facility where purchased animals can become acclimated to new surroundings and visually screened for abnormalities or illness for up to 3 weeks before adding them to the herd. These latter recommendations may help reduce the risk of disease introduction and would be first steps to help secure the herd's health. It would



Johne's disease can cause diarrhea and result in reduced milk yields, lower body weights and beef value, and early culling. It can be detected with a lab test costing about \$5.50 for 10 samples.

TABLE 4. Herd prevalence of different diseases in U.S. dairy cattle (cows, bulls and heifers)

Johne's disease*	Prevalence %
Midwest	60.7
Northeast	26.5
West	8.3
Southeast	4.5
National	22.0 (cows, 5–10%)
BVDV persistent infection†	
Michigan dairy herds	15 (cows, 0.13%)
Bovine leukosis virus (BLV)‡	
Midwest	88.6
Northeast	86.6
West	88.6
Southeast	99.0
National	88.3 (cows, 41%)
Salmonella spp.§	
Midwest	25.6
Northeast	6.7
West	42.9
Southeast	50.0
National	27.5 (cows, 7.3%)
Staph. aureus ¶	
New York/Pennsylvania	9.1 (cows)
Mycoplasma#	
Midwest	2.2 (bulk tanks)
Northeast	2.8
West	9.4
Southeast	6.6
National	7.9
Digital dermatitis (foot warts)**	
Midwest	46–60
Northeast	60.2
West	60–72
Southeast	30.0

* Source: Ott et al. 1999.

† Source: Houe et al. 1995.

‡ Source: Ott et al. 2003.

§ Source: USDA APHIS 2003b.

¶ Source: Wilson et al. 1997.

USDA APHIS 2003a.

** Source: USDA APHIS 1997.

be prudent for producers to develop a protocol for testing risky animals when the herd of origin is unknown or health history is lacking.

Our results indicate that there are opportunities for dairy advisors, herd veterinarians and extension educators to emphasize the risks associated with new herd members and to work with clients on appropriate purchasing and testing strategies. Specifically, they can help producers to: (1) understand the consequences of specific infections, (2) identify and prioritize specific diseases they want to keep out of their herds, (3) assess existing disease conditions in their operation, (4) develop a testing plan for risky animals before or after purchasing (using table 4 and box below), (5) make the best decisions on what to do with infected animals based on available information and (6) assess the operation and facilities for potential within-herd transmission of diseases (CFSPH 2008). Keeping infectious diseases and other cattle conditions out of the herd can save money in the future.

Testing strategies for pre- or post-purchase of dairy herd replacements

All (bulls, cows, heifers)

- Examination for foot warts
- Blood sample for Johne's ELISA test (if over 2 years of age)
- Ear notch or blood sample for BVDV persistent infection test
- Fecal sample for *Salmonella* culture
- Blood sample for bovine leukosis virus (BLV) antigen-capture ELISA

Bulls only

- Breeding soundness investigation, including palpation of scrotal contents and seminal vesicles
- Preputial sample for *Trichomonas* testing

Cows only

- Milk sample for mastitis-pathogen culture

D.A. Moore is Director of Veterinary Medicine Extension, Washington State University; J.M. Adaska is Pathologist, California Animal Health and Food Safety Laboratory; G.E. Higginbotham is Dairy Farm Advisor, UC Cooperative Extension (UCCE), Fresno County; A.R. Castillo is

Dairy Farm Advisor, UCCE Merced County; C. Collar is Dairy Farm Advisor, UCCE Kings County; and W.M. Sischo is Professor of Infectious Disease Epidemiology, Washington State University. This project was funded through a grant from the UC Agriculture and Natural Resources Core Issues Grants Program. The authors thank Marla Hartman for technical assistance.

References

- Buhman M, Dewel G, Griffen D. 2000. Biosecurity Basics for Cattle Operations and Good Management Practices (GMP) for Controlling Infectious Diseases. Nebraska Cooperative Extension G00-1411-A.
- Buttars NK, Young AJ, Bailey D. 2006. Adoption of security measures by dairy farms to address bioterrorist threats in the intermountain United States. *J Dairy Sci* 89:1822-9.
- [CDFA] California Department of Food and Agriculture. 2005. California Dairy Statistics and Trends – 2005 Mid-Year Review. Division of Marketing Services. Sacramento, CA. www.cdfa.ca.gov.
- Casal J, De Manuel A, Mateau E, Martin M. 2007. Biosecurity measures on swine farms in Spain: Perceptions by farmers and their relationship to current on-farm measures. *Prev Vet Med* 82:138-50.
- [CFSPH] Center for Food Safety and Public Health. 2008. Biological Risk Management: Resources for Dairy Facilities. Iowa State University. www.cfsph.iastate.edu/BRM/dairyresources.htm.
- Chi J, VanLeeuwen JA, Weersink A, Keefe GP. 2002. Management factors related to seroprevalences to bovine viral diarrhoea virus, bovine-leukosis virus, *Mycobacterium avium* subspecies paratuberculosis, and *Neospora caninum* in dairy herds in the Canadian maritimes. *Prev Vet Med* 55(1):57-68.
- Collins MM, Gardner IA, Garry FB, et al. 2006. Consensus recommendations on diagnostic testing for the detection of paratuberculosis in cattle in the United States. *J Am Vet Med Assoc* 229(12):1912-9.
- Dalton J, Norell R, Chahine M. 2005a. Biosecurity practices used during dairy herd expansion. *J Dairy Sci* 88(suppl 1):300-1(abstr).
- Dalton J, Norell R, Chahine M. 2005b. Do dairy producers manage dairy bulls to limit biosecurity and infertility risk? *J Dairy Sci* 88(suppl 1):301(abstr).
- D'Angelino JL, Garcia M, Birgel EH. 1998. Productive and reproductive performance in cattle infected with bovine leukosis virus. *J Dairy Res* 65:693-5.
- Faust MA, Kinsel ML, Kirkpatrick MA. 2001. Characterizing biosecurity, health, and culling during dairy herd expansions. *J Dairy Sci* 84:955-65.
- Heald MTS, Waltner-Toews D, Jacobs RM, McNab WB. 1992. The prevalence of anti-bovine leukemia virus antibodies in dairy cows and associations with farm management practices, production and culling in Ontario. *Prev Vet Med* 14:45-55.
- Hoe F, Ruegg PL. 2006. Opinions and practices of Wisconsin dairy producers about biosecurity and animal well-being. *J Dairy Sci* 89:2297-308.
- Houe H, Baker JC, Maes RK, Wuryastuti H. 1995. Prevalence of cattle persistently infected with bovine viral diarrhoea virus in 20 dairy herds in two counties in central Michigan and comparison of prevalence of antibody-positive cattle among herds with different infection and vaccination status. *J Vet Diag Invest* 7:321-6.
- Ott SL, Johnson R, Wells SJ. 2003. Association between bovine leukosis virus seroprevalence and herd-level productivity on US dairy farms. *Prev Vet Med* 61:249-62.
- Ott SL, Wells SJ, Wagner BA. 1999. Herd-level economic losses associated with Johne's disease on US dairy operations. *Prev Vet Med* 40:179-92.
- Pelzer KD. 1997. Economics of bovine leukemia virus infection. *Vet Clin Food Anim* 13(1):129-41.
- Pollari FL, DiGiacomo RF, Evermann JF. 1993. Use of survival analysis to compare cull rates between bovine leukemia virus seropositive and seronegative dairy cows. *Am J Vet Res* 54(9):1400-3.
- Pollari FL, Wangsuphachart VL, DiGiacomo RF, Evermann JF. 1992. Effects of bovine leukemia virus infection on production and reproduction in dairy cattle. *Can J Vet Res* 56(4):289-95.
- Rhodes JK, Pelzer KD, Johnson YJ. 2003. Economic implications of bovine leukemia virus infection in mid-Atlantic dairy herds. *J Am Vet Med Assoc* 223(3):346-52.
- Sandvik T. 2004. Progress of control and prevention programs for bovine viral diarrhoea virus in Europe. *Vet Clin Food Anim* 20(1):151-69.
- Shim EH, Shanks RD, Morin DE. 2004. Milk loss and treatment costs associated with two treatment protocols for clinical mastitis in dairy cows. *J Dairy Sci* 87:2702-8.
- Sockett DC. 1996. Johne's disease eradication and control: Regulatory implications. *Vet Clin Food Anim* 12(2):431-40.
- Tiwari A, VanLeeuwen JA, Dohoo IR, et al. 2007. Production effects of pathogens causing bovine leukosis, bovine viral diarrhoea, paratuberculosis, and neosporosis. *J Dairy Sci* 90:659-69.
- [USDA] US Department of Agriculture. 2008. FSIS Animal Disposition Reporting System. www.fsis.usda.gov/Science/Animal_Disposition_Reporting_System/index.asp (accessed October 2007).
- [USDA APHIS] USDA Animal and Plant Health Inspection Service. 1997. Papillomatous digital dermatitis on U.S. dairy operations. Centers for Epidemiology and Animal Health. Fort Collins, CO. N231.597. 28 p.
- USDA APHIS. 2003a. Mycoplasma in bulk tank milk on U.S. dairies. Centers for Epidemiology and Animal Health. Fort Collins, CO. N395.0503. 3 p.
- USDA APHIS. 2003b. Salmonella and campylobacter on U.S. dairy operations. Centers for Epidemiology and Animal Health. Fort Collins, CO. N401.1203. 3p.
- US Senate. 2006. S 1532, Agroterrorism Prevention Act of 2005. www.govtrack.us.
- Wilson DJ, Gonzalez RN, Das HH. 1997. Bovine mastitis pathogens in New York and Pennsylvania: Prevalence and effects on somatic cell count and milk production. *J Dairy Sci* 80:2592-8.